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CHAIN AND METHODS (57) Abstract The present invention relates to OB protein compose	itions a	TSTABLE, ACTIVE, HUMAN OB PROTEIN WITH ANTIBODY For standard methods. Provided herein are OB protein suspensions which one are useful for the treatment or modulation of weight adiposity lever

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COMPOSITIONS COMPRISING CONJUGATES OF STABLE, ACTIVE, HUMAN OB PROTEIN WITH ANTIBODY FC CHAIN AND METHODS

Field of the Invention

The present invention relates to stable, active human OB protein compositions at high concentrations and at or near physiologic pH. Also provided are related compositions, methods of manufacture and methods of using such compositions.

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Background

Although the molecular basis for obesity is largely unknown, the identification of the "OB gene" and protein encoded ("OB protein") has shed some light on mechanisms the body uses to regulate body fat deposition. Zhang et al., Nature 372: 425-432 (1994); see also, the Correction at Nature 374: 479 (1995). PCT publication No. WO 96/05309, published February 22, 1996, entitled, "Modulators of Body Weight,

- 20 Corresponding Nucleic Acids and Proteins, and
 Diagnostic and Therapeutic Uses Thereof" fully sets
 forth OB protein and related compositions and methods,
 and is herein incorporated by reference. An amino acid
 sequence for human OB protein is set forth at
- WO 96/05309 (herein incorporated by reference) SEQ ID NOS. 4 and 6 (at pages 172 and 174 of that publication), and the first amino acid residue of the mature protein is at position 22 and is a valine residue. The mature protein is 146 residues (or 145 if the glutamine at position 49 is absent, SEQ ID NO. 6).

The OB protein is active in vivo in both ob/ob mutant mice (mice obese due to a defect in the production of the OB gene product) as well as in normal, wild type mice. The biological activity

35 manifests itself in, among other things, weight loss.

See generally, Barinaga, "Obese" Protein Slims Mice,

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Science 269: 475-476 (1995) and Friedman, "The Alphabet of Weight Control," Nature 385: 119-120 (1997). It is known, for instance, that in ob/ob mutant mice, administration of OB protein results in a decrease in serum insulin levels, and serum glucose levels. It is also known that administration of OB protein results in a decrease in body fat. This was observed in both ob/ob mutant mice, as well as non-obese normal mice. Pelleymounter et al., Science 269: 540-543 (1995); Halaas et al., Science 269: 543-546 (1995). See also, 10 Campfield et al., Science 269: 546-549 (1995) (Peripheral and central administration of microgram doses of OB protein reduced food intake and body weight of ob/ob and diet-induced obese mice but not in db/db obese mice.) In none of these reports have toxicities 15 been observed, even at the highest doses.

For preparation of a pharmaceutical composition for injection in humans, it has been observed that the human amino acid sequence is insoluble at physiologic pH at relatively high concentrations, such as above about 2 mg active protein/milliliter of liquid. Dosages in the milligram protein per kilogram body weight range, such as .5 or 1.0 mg/kg/day or below, are desirable for injection of 25 therapeutically effective amounts into larger mammals, such as humans. An increase in protein concentration is necessary to avoid injection of large volumes, which can be uncomfortable or possibly painful to the patient.

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With the advances in recombinant DNA technology, the availability of recombinant proteins for therapeutic use has engendered advances in protein formulation. A review article describing protein

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modification and fusion proteins is Francis, Focus on Growth Factors 3:4-10 (1992).

One such modification is the use of the Fc region of immunoglobulins. Antibodies comprise two 5 functionally independent parts, a variable domain known as "Fab", which binds antigen, and a constant domain, known as "Fc" which provides the link to effector functions such as complement or phagocytic cells. The Fc portion of an immunoglobulin has a long plasma halflife, whereas the Fab is short-lived. Capon et al., Nature 337: 525-531 (1989).

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Therapeutic proteins have been constructed using the Fc domain to provide longer half-life or to incorporate functions such as Fc receptor binding, protein A binding, complement fixation and placental transfer which all reside in the Fc proteins of immunoglobulins. Id. For example, the Fc region of an IgG1 antibody has been fused to the N-terminal end of CD30-L, a molecule which binds CD30 receptors expressed on Hodgkin's Disease tumor cells, anaplastic lymphoma 20 cells, T-cell leukemia cells and other malignant cell types. See, U.S. Patent No. 5,480,981. IL-10, an antiinflammatory and anti-rejection agent has been fused to murine Fcy2a in order to increase the cytokine's short 25 circulating half-life. Zheng, X. et al., The Journal of Immunology, 154: 5590-5600 (1995). Studies have also evaluated the use of tumor necrosis factor receptor linked with the Fc protein of human IgG1 to treat patients with septic shock. Fisher, C. et al., 30 N. Engl. J. Med., 334: 1697-1702 (1996); Van Zee, K. et al., The Journal of Immunology, 156: 2221-2230 (1996). Fc has also been fused with CD4 receptor to produce a therapeutic protein for treatment of AIDS. See, Capon et al., Nature, 337:525-531 (1989). In addition, the

N-terminus of interleukin 2 has also been fused to the Fc portion of IgG1 or IgG3 to overcome the short half life of interleukin 2 and its systemic toxicity. See, Harvill et al., Immunotechnology, 1: 95-105 (1995).

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been reported. But, those conditions which are applicable to insulin are not predictive of those conditions which may be applicable to any other protein, including OB protein. Insulin is a fairly small protein, having particular physical and chemical characteristics; these characteristics are important to determining conditions for formulation. Brange, Galenics of Insulin, Springer-Verlag 1987 describes insulin suspensions (p.36); see also, Schlichtkrull et al. Insulin Preparations with Prolonged Effect, pp. 729-777 In: Hassellblatt et al., Handbook of Experimental Pharmacology New Series, Vol. XXXII-1/2 Springer-Verlag Berlin, Heidelburg, New York (1975).

To date, there have been no reports of stable preparations of human OB protein at concentrations of at least about 2 mg/ml at physiologic pH, and further, no reports of stable concentrations of active human OB protein at least about 50 mg/ml or above. Moreover, a frozen or lyophilized form may be used to improve shelf-stability but, are less desirable than the readyto-use suspension forms described here. A frozen form requires storage at a constant frozen temperature which may not be possible due to defrost cycles of consumergrade refrigerators and freezers. Furthermore, a lyophilized form must be diluted and mixed which is inconvenient and may compromise patient compliance. From a producer's point of view, manufacturing, storing and shipping of frozen liquid is expensive and requires far more supervision than distribution of a ready to

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use formulation. Also, manufacture or otherwise making available a suitable diluent for a lyophilized formulation is more costly and less efficient than not requiring such diluent. There exists a need for 5 concentrated forms of human pharmaceutical compositions containing active OB protein which can be delivered by injection at low volumes. The present invention fulfills these requirements.

Summary of the Invention 10

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The present invention stems from the observation that certain suspension formulations allow the preparation of stable OB protein in concentrations of at least about 2 mg/ml at physiological pH. As used herein, the term, "physiological pH" refers to a pH of about 6.0 to about 8.0. Use of such compositions allow for delivery of relatively low volumes of OB protein therapeutic. As further disclosed herein, the use of precipitating agents allows for the preparation of suspensions of human OB protein having the 20 characteristics of:

Improved stability at physiological pH as compared to the same human OB protein in solution form. Demonstrated below in the working examples are human OB protein suspensions at concentrations of 10 mg/ml or higher and at pH 7, which, when compared to the same concentration in solution at the highest pH which allows solubility (pH 4 for human OB protein in its native form, which is not even physiologic pH) have better HPLC (high pressure liquid chromatography) profiles. Also demonstrated below is a Fc-OB fusion protein suspension at a concentration of 5 mg/ml at about pH 7 which shows less degradation products upon

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storage than a comparable Fc-OB fusion protein solution.

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2. Sustained injection time-release profile as compared to the same human OB protein in solution form. The working examples below demonstrate a sustained-release effect using the present highly concentrated human OB protein suspensions. Such sustained release effect is advantageous in that the activity of material is maintained over a relatively longer period of time, and thus there is a relatively higher potency, and the need for fewer injections as compared with human OB protein in solution.

Thus, one object of the present invention is a stable preparation of active human OB protein at physiologic pH, having a concentration of at least about 2 mg protein/ml. For instance, a stable preparation of active human OB protein at a concentration of at least 2.0 mg/ml and a pH of 7.0 is provided. The upper limit of concentration is that suspension form which is available for administration to a human, and, as described below, working examples have provided a concentration of as high as 100 mg/ml at physiologic pH (e.g., between pH 6.0 and pH 8.0).

In another aspect, the present invention relates to a stable preparation of active human OB protein within the pH range of about 6.0 to about 8.0 having a concentration of at least about 10 mg/ml.

In still another aspect, the present invention relates to a stable preparation of active human OB protein derivatized by the attachment of an Fc region of an immunoglobulin at a concentration of at least about 0.5 mg/ml and a pH of about 6.0 to about 8.0. More particularly, stable preparations of active

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Fc-OB fusion protein at a concentration of 5 mg/ml to 50 mg/ml at about pH 7.5 are provided.

In yet other aspects, the present invention provides formulations for stable, active human OB

protein at a concentration of 2 mg protein/ml or above, at pH 6.5 to pH 7.5. More particularly, formulations for stable, active human OB protein at a concentration of 20 mg/ml to 100 mg/ml at pH 7.0 are provided.

In another aspect, the present invention provides formulations for stable, active human OB protein at a concentration of 10 mg/ml or above at a pH of 5.0 to 8.0.

In still another aspect, the present invention provides formulations for stable, active human OB protein derivatized by the attachment of an Fc region of an immunoglobulin at a concentration of 0.5 mg/ml or above and a pH of about 6.0 to about 8.0. More particularly, formulations for stable, active Fc-OB fusion protein at a concentration of 5 mg/ml to 50 mg/ml at about pH 7.5 are provided.

Yet other aspects of the present invention include pharmaceutical compositions of the above, methods for manufacturing such compositions, and methods of treatment using present compositions, and methods of manufacturing medicaments containing the present compositions for such treatment.

Brief Description of the Drawings

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FIGURES 1A and 1B are dose response curves in 30 mice for (1A) the present human OB suspension of Example 1 and (1B) a control human OB protein solution as described in Example 1.

FIGURE 2 is a graph illustrating OB serum levels in dogs for the present OB protein suspension

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and for a control human OB protein solution as described in Example 1.

FIGURE 3 is a reversed phase high pressure liquid chromatography (RP-HPLC) tracing of human OB 5 protein formulations at 37°C for seven weeks: human OB protein zinc suspension of Example 1 is the middle tracing, the human OB protein solution control of Example 1 is the top line and a human OB protein suspension kept at -80°C for 56 days is the bottom line.

FIGURE 4 is a RP-HPLC tracing of human OB protein formulations at 19°C for 56 days: human OB solution of Example 1 is the top line, the human OB crystalline suspension of Example 2 is the middle line, and the human OB crystalline suspension of Example 2 at -80°C for 56 days is the bottom line.

FIGURES 5A-5C are the DNA sequence (SEQ ID NO: 1) and amino acid sequence (SEQ ID NO: 2) of a human metFc-OB protein.

FIGURES 6A-6C are the DNA sequence (SEQ ID 20 NO: 3) and amino acid sequence (SEQ ID NO: 4) of a human metFc-OB protein variant.

FIGURE 7 is a graph depicting the rate of formation of iso asp at asp 108 (asp 335 using the numbering according to SEQ ID NO: 4), as determined by reverse phase HPLC, for recombinant methionyl human Fc-OB protein.

<u>Detailed Description</u>

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The present stable, active OB protein 30 compositions are generally classified as suspensions, in that the protein moiety is precipitated and suspended in a liquid moiety. The compositions contain an active OB protein moiety, a precipitating agent, a

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pH modulating agent, and a liquid carrier. The present OB proteins are either amorphous or in crystalline form.

Preferably, for use as a therapeutic or 5 cosmetic composition in humans, OB protein with the amino acid sequence of native human OB protein (see Zhang et al., Nature, supra), optionally with an N-terminal methionyl residue incident to bacterial expression, is used. See, PCT Publication WO 96/05309, herein incorporated by reference, for recombinant DNA means to prepare the present OB proteins which may be used. One may make changes in selected amino acids so long as such changes preserve the overall folding or activity of the protein. Table 1, below sets forth conserved amino acid substitutions which may be used, 15 in terms of particular characteristics (basic, acidic, polar, hydrophobic, aromatic, and size (small)). See, in general, Ford et al., Protein Expression and Purification 2: 95-107, 1991, which is herein incorporated by reference. Small amino terminal 20 extensions, such as an amino-terminal methionine residue, a small linker peptide of up to about 20-25 residues, or a small extension that facilitates purification, such as a poly-histidine tract, an antigenic epitope or a binding domain, may also be 25 present.

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Table 1
Conservative Amino Acid Substitutions

Basic:	arginine lysine histidine
Acidic:	glutamic acid aspartic acid
Polar:	glutamine asparagine
Hydrophobic:	leucine isoleucine valine
Aromatic:	phenylalanine tryptophan tyrosine
Small:	glycine alanine serine threonine methionine

Generally, human OB proteins which will display increased stability in the present suspensions will be those which, upon exposure to physiologic pH, have an exposed hydrophobic region when in solution. Additionally, human OB proteins derivatized by the attachment of an Fc region of an immunoglobulin to the OB protein moiety display increased stability in the present suspensions.

Generally, an Fc region of an immunoglobulin may be genetically or chemically fused to a human OB protein. Preferably, the Fc region is fused at the N-terminus of the OB protein. See copending U.S. application Serial No. 08/770,973, filed December 20, 1996, herein incorporated by reference, for preferred Fc-OB fusion proteins.

20 Preferably, an Fc region with the amino acid sequence of the human immunoglobulin IgG-1 heavy chain (see Ellison, J.W. et al., Nucleic Acids Res. 10: 4071-

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4079 (1982)) is used. A preferred Fc region is set forth in SEQ ID NO: 2 (see Figure 5). The recombinant Fc-OB sequence of SEQ ID NO: 2 is a 378 amino acid Fc-OB protein (not counting the methionine residue). The first amino acid residue of the Fc-OB protein in Figure 5, glutamic acid, is referred to as +1 with the methionine at the -1 position. Variants or analogs of the Fc portion may be constructed by, for example, making various substitutions of amino acid residues or base pairs.

Cysteine residues can be deleted or replaced with other amino acids to prevent formation of disulfide crosslinks of the Fc sequences. In particular, amino acid at position 5 of SEQ ID NO: 2 is a cysteine residue. One may remove the cysteine residue at position 5 or substitute it with one or more amino acids. For example, an alanine residue may be substituted for the cysteine residue at position 5 resulting in a variant amino acid sequence. Likewise, the cysteine at position 5 of SEQ ID NO: 2 may be substituted with a serine or other amino acid residue or deleted.

A variant or analog may also be prepared by deletion of the amino acids at positions 1, 2, 3, 4 and 5, resulting in a 373 amino acid Fc-OB protein (not counting the methionine residue). This sequence is set forth in SEQ ID NO:4 (see Figure 6). Substitutions at these positions can also be made and are within the scope of this invention.

Modifications may also be made to introduce four amino acid substitutions to ablate the Fc receptor binding site and the complement (Clq) binding site.

According to the numbering of SEQ ID NO: 4, these variant modifications include 1 ucine at position 15

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substituted with glutamic acid, glutamic acid at position 98 substituted with alanine, and lysines at positions 100 and 102 substituted with alanines.

Likewise, one or more tyrosine residues can 5 be replaced by phenylalanine residues as well. As described above, one may make changes in selected amino acids as long as such changes preserve the overall folding or activity of the fusion protein.

Furthermore, the Fc region may be also linked to the human OB protein of the Fc-OB fusion protein by "linker" moieties whether chemical or amino acids of varying lengths. Such chemical linkers are well known in the art. Amino acid linker sequences can include but are not limited to:

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4 -	(a)	ala, ala, ala;
15	(a)	ala, ala, ala,
	(b)	ala, ala, ala, ala;
	(c)	ala, ala, ala, ala;
	(b)	gly, gly;
	(e)	gly, gly, gly;
20	(f)	gly, gly, gly, gly;
	(g)	gly, gly, gly, gly, gly, gly, gly;
	(h)	gly-pro-gly;
	(i)	gly, gly, pro, gly, gly; and
	(j)	any combination of subparts (a)
25	through (i).	

The present precipitating agents may be a salt having a cationic component, and may be selected from among calcium, magnesium, zinc, sodium, iron, cobalt, manganese, potassium and nickel. Preferably, the salt will be compatible for use in a pharmaceutical composition.

Alternatively precipitating agents may be selected from among agents which are pharmaceutically acceptable yet known to precipitate protein, such as

polyethylene glycols, or other water soluble polymers as set forth in the next paragraphs. A useful precipitating agent will induce precipitation of OB protein at neutral pH but is reversible or 5 redissolveable upon dilution with physiologically compatible solvents. Without an appropriate precipitating agent, OB protein precipitates at neutral pH to a form that is not reversible by dilution with physiologically compatible solvents.

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The pH range is preferably from about pH 4.0 to about pH 8.0, and more preferably from about 6.5 to about 7.5. The most preferable pH for a pharmaceutical composition is that in which the OB protein used may retain its maximum biological activity at the selected protein concentration. At non-physiologic pH, the 15 present OB protein suspensions may also have advantages. At pH below 5.0, the present OB protein suspensions may be more stable (in terms of shelf-life) than equal concentrations of OB protein in solution at equal pH. For example at pH 4.0, and a concentration of 20 50 mg/ml, the present suspensions may have greater biological activity upon in vivo administration than would the equivalent solution.

The buffer may be selected from among those 25 which attain the desired pH while not altering the precipitating characteristics of the composition. Preferably, buffers will be acceptable for a pharmaceutical formulation. Tris, MES, and PIPES are acceptable for both the amorphous and crystalline forms. Phosphate is a preferred buffer for the 30 crystalline forms.

The final suspension will preferably have a concentration of 5 mg/ml to 100 mg/ml for ease of therapeutic administration.

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Methods of Use

Therapeutic. Therapeutic uses include weight modulation, the treatment or prevention of diabetes, blood lipid reduction (and treatment of related conditions), increasing lean body mass and increasing insulin sensitivity. In addition, the present compositions may be used for manufacture of one or more medicaments for treatment or amelioration of the above conditions. Methods of administration will typically be by injection, although other means, such as pulmonary delivery may be used. See PCT WO 96/05309, incorporated by reference at page 83 et seq., for example. The present suspensions may be spray-dried into particles having an average size of less than 10 microns, or more preferably, 0.5 to 5 microns.

Weight Modulation. The present compositions and methods may be used for weight reduction. Viewed another way, the present compositions may be used for maintenance of a desired weight or level of adiposity. As has been demonstrated in murine models (see supra), administration of the present OB protein results in weight loss. The body mass lost is primarily of adipose tissue, or fat. Such weight loss can be 25 associated with the treatment of concomitant conditions, such as those below, and therefore constitute a therapeutic application. In addition, cosmetic uses are provided herein if weight modulation is solely for improvement in appearance.

Treatment of Diabetes. The present compositions and methods may be used in the prevention or treatment of Type II diabetes. As Type II diabetes can be correlated with obesity, use of the present invention to reduce weight (or maintain a desired

weight, or reduce or maintain an adiposity level) can also alleviate or prevent the development of diabetes. Moreover, even in the absence of dosages sufficient to result in weight loss, the present compositions may be used to prevent or ameliorate diabetes.

Blood Lipid Modulation. The present compositions and methods may be used in the modulation of blood lipid levels. Ideally, in situations where solely reduction in blood lipid levels is desired, or where maintenance of blood lipid levels is desired, the dosage will be insufficient to result in weight loss. Thus, during an initial course of therapy of an obese patient, dosages may be administered whereby weight loss and concomitant blood lipid level lowering is achieved. Once sufficient weight loss is achieved, a dosage sufficient to prevent re-gaining weight, yet sufficient to maintain desired blood lipid levels, or other conditions as set forth herein, for example, be administered. These dosages can be determined empirically, as the effects of OB protein are reversible. E.g., Campfield et al., Science 269: 546-549 (1995) at 547. Thus, if a dosage resulting in weight loss is observed when weight loss is not desired, one would administer a lower dose in order to achieve the desired blood lipid levels, yet maintain the desired weight. See, e.g., PCT Publication WO 97/06816 herein incorporated by reference.

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Increasing Lean Mass or Insulin Sensitivity.

Ideally, in situations where solely an increase in lean
body mass is desired, the dosage will be insufficient
to result in weight loss. Thus, during an initial
course of therapy of an obese person, dosages may be
administered whereby weight loss and concomitant fat
tissue decrease/lean mass increase is achieved. Once

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sufficient weight loss is achieved, a dosage sufficient to prevent regaining weight, yet sufficient to maintain desired lean mass increase (or prevention of lean mass depletion) may be administered. For increasing an individual's sensitivity to insulin, similar dosage considerations may be taken into account. Lean mass increase without weight loss may be achieved sufficient to decrease the amount of insulin (or, potentially, amylin, amylin antagonists or agonists, or thiazolidinediones, or other potential diabetes 10 treating drugs) an individual would be administered for the treatment of diabetes. For increasing overall strength, there may be similar dosage considerations. Lean mass increase with concomitant increase in overall strength may be achieved with doses insufficient to 15 result in weight loss. Other benefits, such as an increase in red blood cells (and oxygenation in the blood) and a decrease in bone resorption or osteoporosis may also be achieved in the absence of weight loss. See, e.g., PCT Publication No. WO 20 97/18833 herein incorporated by reference.

Combination Therapies. The present compositions and methods may be used in conjunction with other therapies, such as altered diet and exercise. Other medicaments, such as those useful for the treatment of diabetes (e.g., insulin and possibly amylin, antagonists or agonists thereof, thiazolidinediones (see, e.g., PCT Publication No. WO 98/08512 herein incorporated by reference), or other potential diabetes treating drugs), cholesterol and blood pressure lowering medicaments (such as those which reduce blood lipid levels or other cardiovascular medicaments), activity increasing medicaments (e.g., amphetamines), diuretics (for liquid elimination), and

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appetite suppressants (such as agents which act on neuropeptide γ receptors or serotonin reuptake inhibitors). Such administration may be simultaneous or may be in seriatim. In addition, the present methods may be used in conjunction with surgical procedures, such as cosmetic surgeries designed to alter the overall appearance of a body (e.g., liposuction or laser surgeries designed to reduce body mass, or implant surgeries designed to increase the appearance of body mass). The health benefits of cardiac surgeries, such as bypass surgeries or other surgeries designed to relieve a deleterious condition caused by blockage of blood vessels by fatty deposits, such as arterial plaque, may be increased with concomitant use of the present compositions and methods. Methods to eliminate gall stones, such as ultrasonic or laser methods, may also be used either prior to, during or after a course of the present therapeutic methods. Furthermore, the present methods may be used as an adjunct to surgeries or therapies for broken bones, damaged muscle, or other therapies which would be improved by an increase in lean tissue mass.

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The following examples are offered to more fully illustrate the invention, but are not to be construed as limiting the scope thereof. Example 1 sets forth preparation of an amorphous (as opposed to a crystalline, as infra) human OB protein suspension at a concentration of 100 mg/ml at pH 7.0. Example 2 sets forth preparation of a crystalline OB protein suspension. Example 3 demonstrates an improved dose response for OB protein suspensions as compared to OB protein solutions. Example 4 demonstrates the delayed time action profile of the present suspensions in a dog model. Example 5 sets forth preparation of an

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amorphous Fc-OB protein suspension. Examples 6 and 7 demonstrate the improved stability of the present suspensions.

EXAMPLE 1: Preparation of an amorphous OB protein suspension.

This example illustrates preparation of one of the human OB protein suspensions of the present invention. An amorphous human OB protein suspension was prepared by precipitation with zinc salt. At a final pH of 6.0 to pH 8.0, a concentration of 100 mg protein/ml liquid has been obtained. Also set forth is a control composition, of pH 4.0, for a human OB protein solution.

Composition: 15

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Protein moiety: recombinant methionyl human OB protein ("rmetHu-leptin") as set forth in SEQ ID NO. 4 of PCT Publication WO 96/05309, beginning with amino acid number 22 (Val) and ending with amino acid number 167, having at its N-terminus a methionyl residue:

Precipitant: Zinc chloride Buffer: Tris, MES and Pipes Final pH: 6.0-8.0

Preparation protocol: Recombinant methionyl human OB protein ("rmetHu-leptin") solution was concentrated to about 40 mg/ml in water for injection, acidified to pH 3.0, with HCl. Zinc chloride was added and the suspension was formed by adjusting the pH to 30 near neutrality (approximately pH 7.0) by adding an appropriate buffer. Tris, MES and PIPES buffer have been successfully used. The final conditions were typically 10 to 15 mM buffer, 20 to 1000 µM Zinc, pH 6.0 to 8.0 and 10 mg/ml rmetHu-leptin. The

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suspensions have been concentrated by allowing the particles to settle at 4°C for several hours and removing the supernatant. This procedure may be repeated several times until the maximum concentration is obtained, and has been used to obtain about 100 mg/ml suspensions.

Control Composition: Recombinant methionyl human OB protein (as above) solution at 20 mg/ml pH 4.0 containing 10 mM acetate and 5% w/v sorbitol was used as a control composition.

EXAMPLE 2: Preparation of a crystalline OB protein suspension.

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This example illustrates the preparation of a crystalline OB protein suspension of the present invention.

Protein moiety: recombinant methionyl human OB protein as in Example 1, above, was used.

Procedure: rmetHu-leptin at a concentration of 15 mg/ml in 1 mM HCl was mixed in a 1:1 ratio with 20 4 M NaCl, 100 mM Tris, pH 8.5, 2% v/v ethanol, at 4°C. Crystals spontaneously formed by slowly adjusting the temperature over several hours to between 14°C and 25°C and maintaining that temperature for at least 2 hours depending on the duration of warming to the final 25 temperature. The "mother liquor" (i.e., the liquid in which the crystals were grown) was replaced with a more suitable solvent for injection by harvesting the crystals by centrifugation and resuspension in an appropriate crystal stabilizing solvent. A suitable 30 replacement solvent is 20-25% polyethylene glycol (having a molecular weight of about 4000 daltons to about 20,000 daltons, the term "about" meaning that the approximate average of molecular weights for commercial

PEG' preparations), appropriate neutral pH buffer, preferably about pH 6.0 to about pH 8.0, and preferably 10 mM phosphate buffer pH 6.0 to pH 7.5, and 2% ethanol v/v. In a typical preparation, some residual salt, usually less than 0.25 M, may remain.

<u>EXAMPLE 3</u>: Improved dose response for OB protein suspensions as compared to OB protein solutions.

This example demonstrates that the present OB protein suspensions are more efficacious than OB 10 protein in solution. Normal lean mice were given daily injections, for 5 days, of either the present suspension, at 1, 10 and 50 mg of protein per kg of body weight, or OB protein solution at the same dose. The mice given the suspension lost more weight per unit 15 of mass of OB protein given than did mice given equal doses of the solution formulation. This is illustrated in FIGURE 1A and FIGURE 1B. FIGURE 1A shows the percent weight change when the suspension of Example 1 was given. FIGURE 1B shows the percent weight change 20 when the control solution of Example 1 was given.

This illustrates that the present suspension at the same dose is more efficacious than that given in solution form. While not wishing to be bound by theory, this may be due to slower absorption rate of the suspension as compared to the solution. The suspension must dissolve before it enters the blood, so this sustained release effect results in higher efficacy. On a mass basis, less protein in suspension needs to be administered than solution. Further, in solution there is no difference between the 10 and 50 mg/kg dose at day 5 (the last day of dosing) (See Table 2, below). The suspension gives a more definitive dose response curve than does the solution formulation.

25

TABLE 2
PERCENT WEIGHT CHANGE FROM DAY 0

Day	6
Placebo	1.4
Suspension 1 mg/kg	-1.7
Suspension 10 mg/kg	-5.9
Suspension 50 mg/kg	-9.5
Solution 1 mg/kg	7
Solution 10 mg/kg	-3.0
Solution 50 mg/kg	-3.5

5 The date is an average for 5 mice per treatment.

Methods:

Animals: Normal CD-1 mice were used.

Base weight: About 20 grams.

10 Administration: Animals were injected SC each day

in the same location for 5 days.

Handling: Animals were group housed, and fed ad

libitum.

Compositions:

Solution: The rmetHu-leptin solution of Example 1 was

used, at a concentration of 20 mg/ml.

Suspension: The rmetHu-leptin suspension of Example 1

was used, at pH 7.0, 10 mM MES buffer,

500 mM Zn, 20 mg/ml.

20 PBS: Phosphate buffered saline was used as a placebo.

(Dose responses for controls using suspension liquid or solution liquid alone, with no protein, were similar to PBS, data not shown).

25 EXAMPLE 4: OB protein serum levels in dogs.

- 22 -

This Example demonstrates the delayed time action profile of the present suspensions.

Methods: Beagle dogs were administered suspension or

solution rmetHu-leptin. Serum was drawn, and OB

protein levels were measured at the times illustrated in FIGURE 2.

TABLE 3

Time after injection (hours)	Serum concentratio n Solution	Serum concentration Suspension (crystal)	Serum concentration Suspension (zinc)
0	0	0	0
.5	532	9.4	50.5
1	1047	12.9	130.4
2	1706	51	298.5
4	1480	87	323.5
8	742	298	247.7
12	301	426	241.1
16	109	389	209.6
24	5	90	57.7

10

Compositions used:

Solution: The solution according to Example 1 was used, at a dose of 5 mg/kg/day.

Suspension: The suspension according to Example 2

15 was used. See Table.

Animals: The data presented in Table 3 is for an average of 3 animals. Animals were normal beagle dogs.

Handling: Animals were housed individually, and fed <u>ad</u>

20 <u>libitum</u>. Good animal handling practices were obeyed.

- 23 -

An antibody assay was used as in Hotta et al, Assav: J. Biol. Chem <u>271</u>: 255327-25331 (1996). Results: As can be seen from FIGURE 2, the solution concentration peaks at 1 to 2 hours after 5 administration of protein, whereas the suspension concentration peaks much later, after 10-12 hours. This demonstrates that the suspension maintains a minimum effective dose for a longer time period.

EXAMPLE 5: Preparation of an amorphous Fc-OB protein 10 suspension.

This example illustrates preparation of one of the human Fc-OB protein suspensions of the present invention. An amorphous human Fc-OB protein suspension was prepared by precipitation with zinc salt. Also set forth is a control composition, of pH 7.5, for a human Fc-OB protein solution.

Composition:

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Protein moiety: recombinant methionyl human 20 Fc-OB protein ("rmetHu-Fc-leptin") as set forth in SEQ ID NO: 4, a 373 amino acid fusion protein, the Fc portion of the protein being amino acids 1-227 and the human OB protein portion of the protein being amino acids 228-373, having at its N-terminus a methionyl residue:

> Precipitant: Zinc chloride Buffer: 100mM Tris Final pH: 7.5

Preparation protocol: A rmetHu-Fc-leptin 30 solution was concentrated to about 5 mg/ml in 100 mM Tris buffer, pH 7.5. Zinc chloride was added to form the suspension.

Control Composition: A rmetHu-Fc-leptin solution (as above) at 5 mg/ml, pH 7.5 containing 100 mM Tris was used as a control composition.

5 EXAMPLE 6: Stability of the present human OB protein suspensions.

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This example demonstrates that both the amorphous and the crystalline forms of the present suspensions are more stable under accelerated stability assay conditions than material in solution.

FIGURE 3 illustrates an RP-HPLC tracing comparing the zinc amorphous form of the present suspension (as in Example 1) with the solution form (also as in Example 1), at 37°C for a period of 7 weeks. As can be seen, the present suspension (middle line) has fewer peaks (indicating fewer breakdown products) than the solution form (top line). As a comparison, the bottom line presents the zinc amorphous form which had been stored for 7 weeks at -80°C.

FIGURE 4 illustrates an RP-HPLC tracing comparing the crystalline suspension form of Example 2 with the solution form (of Example 1). The materials were stored for 56 days at 19°C. (Storage at 37°C was not a relevant condition, as the crystalline form loses its crystalline structure at 37°C.) FIGURE 4 shows that there are more peaks, indicating more degradation, for the solution form, (main peak = 86%) than for the suspension form (main peak = 94%). The control at the bottom line was the crystalline form stored for 56 days at -80°C. Similar results were observed for 4°C, although degradation occurred more slowly.

The main degradation product appeared to be the aspartate at amino acid position 108 (according to SEQ ID NO: 4 in PCT WO 96/05309, using the "Val"

- 25 -

residue as position number 1). One may empirically select a more stable chemical moiety at this position, such as another amino acid, to improve stability of the molecule. Overall, the suspensions of the present invention are more stable than OB protein in solution.

EXAMPLE 7: Stability of the present human Fc-OB protein suspensions.

This example demonstrates that the amorphous form of the present Fc-OB protein suspensions is more 10 stable under accelerated stability assay conditions than material in solution.

15

The zinc amorphous form of the present suspension (as in Example 5) and the solution form (also as in Example 5) were put on stability studies by storing both the suspension and solution forms at 4°C, 29°C and 37°C for a period of 2 weeks. As shown in FIGURE 7, the Fc-leptin suspensions stored at 29°C and 37°C show a lower percentage of degradation products 20 related to the aspartate at amino acid position 108 of the OB protein than the Fc-leptin solutions stored at the same temperatures, respectively.

While the present invention has been 25 described in terms of preferred embodiments, it is understood that variations and modifications will occur to those skilled in the art. Therefore, it is intended that the appended claims cover all such equivalent 30 variations which come within the scope of the invention as claimed.

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CLAIMS

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1. A human OB protein suspension having a pH
of about 6.0 to about 8.0 and a concentration of at
least 0.5 mg/ml wherein the OB protein is derivatized
by the attachment of an Fc region of an immunoglobulin
to the N-terminus of the OB protein moiety.

- 2. A human OB protein suspension of claim 1

 wherein the Fc portion of said human Fc-OB protein is selected from the group consisting of:
 - (a) the Fc amino acid sequences as set forth in SEQ ID NOS: 2 and 4;
 - (b) the amino acid sequence of subpart (a) having a different amino acid substituted or deleted in one or more of the following positions (using the numbering according to SEQ ID NO: 2):
 - (i) one or more cysteine residuesreplaced by an alanine or serine residue;
- 20 (ii) one or more tyrosine residues replaced by a phenylalanine residue;
 - (iii) the amino acid at position 5
 replaced with an alanine;
 - (iv) the amino acid at position 20
 replaced with glutamic acid;
 - (v) the amino acid at position 103 replaced with an alanine;
 - (vi) the amino acid at position 105
 replaced with an alanine;
- 30 (vii) the amino acid at position 107 replaced with an alanine;
 - (viii) the amino acids at positions
 1, 2, 3, 4, and 5 deleted;

- 27 -

(ix) one or more residues replaced or deleted to ablate the Fc receptor binding site;

(x) one or more residues substituted or deleted to ablate the complement (Clq) binding site; and

(xi) a combination of subparts i-x; and

5

- (c) the amino acid sequence of subparts (a) or (b) having a methionyl residue at the N
 terminus.
 - 3. A human OB protein suspension of claim 1 wherein said concentration is at least 5 mg/ml.
- 4. A human OB protein suspension of claim 1 wherein said concentration is between 5 mg/ml and 50 mg/ml.
- 5. A human OB protein suspension of claim 1 20 wherein said pH is pH 7.0.
 - 6. A human OB protein suspension of claim 2 wherein said human OB protein is rmetHu-Fc-leptin.
- 7. A human OB protein suspension of claim 1 containing a pharmaceutically acceptable precipitating agent.
- 8. A human OB protein suspension of claim 1
 30 containing a precipitating agent selected from a salt and a water soluble polymer.
 - 9. A human OB protein suspension of claim 1 containing a precipitating agent selected from among

- 28 -

calcium, magnesium, zinc, sodium, iron, cobalt, manganese, potassium and nickel.

- 10. A method of treating an individual for a condition by administering an effective dose of a human OB suspension according to claim 1, said condition selected from among:
 - (a) weight modulation,
 - (b) adiposity modulation,
- 10 (c) diabetes,

20

- (d) blood lipid level modulation,
- (e) increase in lean mass, and
- (f) increase in insulin sensitivity.
- 15 11. A method of preparing a human OB protein suspension comprising:

combining, under suitable conditions, a precipitating agent and a human OB protein in solution, wherein the OB protein is derivatized by the attachment of an Fc region of an immunoglobulin to the N-terminus of the OB protein moiety; allowing said human OB protein to precipitate;

collecting said precipitated human OB protein; and optionally, resuspending said human OB protein in a diluent.

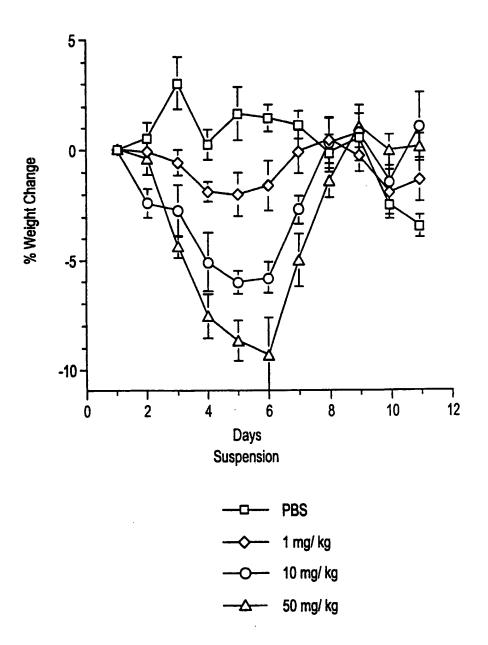
30 12. A method of claim 11 wherein said human OB protein is resuspended in a diluent having a pH of between 6.0 and 8.0.

- 29 -

13. A method of claim 11 wherein said human OB protein is rmetHu-Fc-leptin.

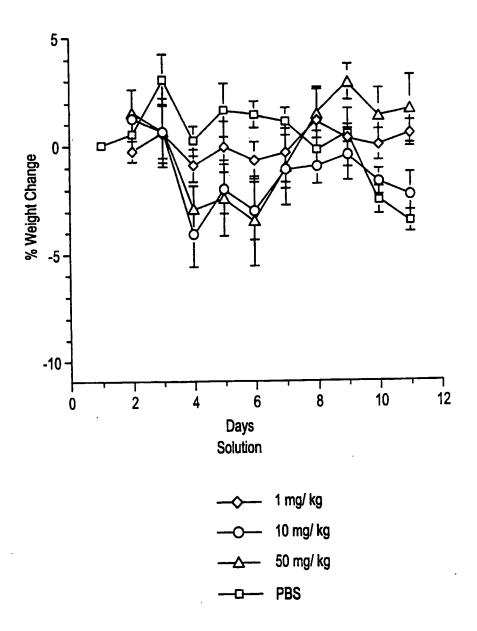
1/12

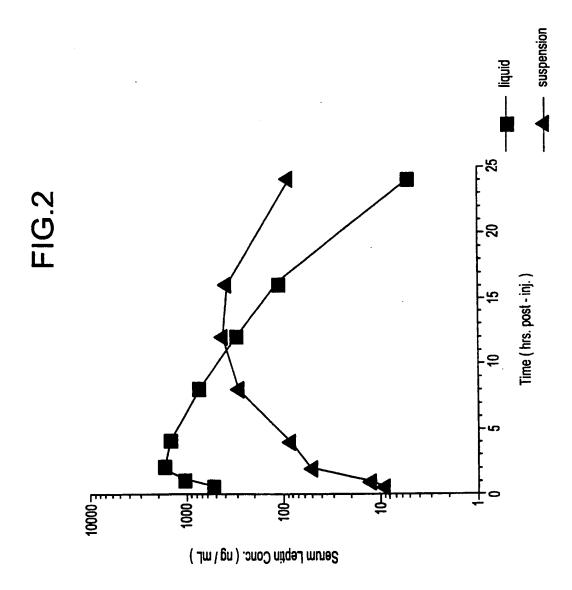
FIG. 1A



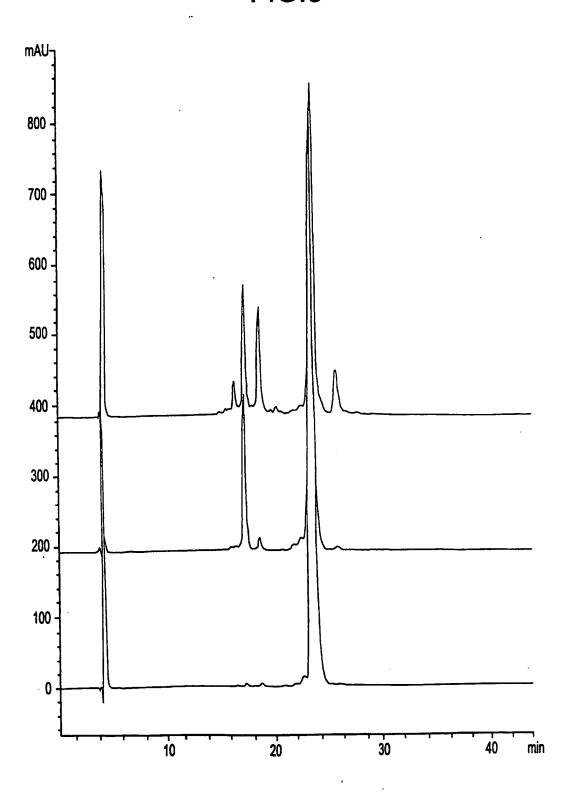
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FIG. 1B





4 / 1 2 FIG.3



5 / 1 2 FIG.4

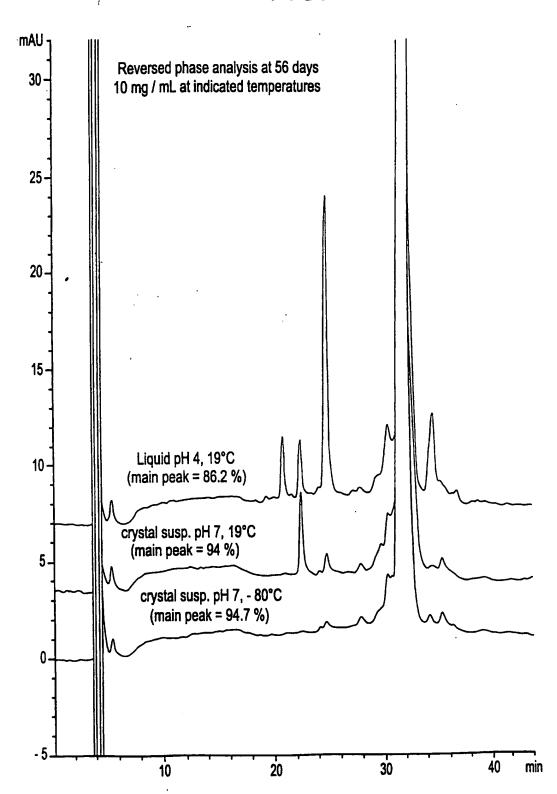


FIG.5A

Recombinant human metFc-OB (double stranded DNA) and amino acid sequence (SEO ID NOS: 1 and 2).

5	1							TGTG			+			-+-			+			rgaa + actt	60
10			M M					C I													•
	61							-+			+			-+-			+		• • •	GATC + CTAG	120
15				G				V F													•
20	121	AGG	GCC	CTGG	+	CTC	CAG	TGT	ACGC	ACC	.+	CCT	GCA	CTC	GGT	GCT	+	GGG	ACT(GGTC + CCAG	180
		s	R	T	.P	E	V	T (: V	V	V	D	V	S	H	E	D	P	E	V	•
25	181							-+			. +			-+-			+	• • •		GGAG + CCTC	240
								ם מ													-
30	241				. +			-+-			-+		• • •	-+-			+			CTGG + GACC	300
		E	Q	Y	N	s	T	Y	r v	v	s	V	L	T	V	L	Н	Q	D	W	•
35	301				. 4	. .		-+-			- + - •	. 		-+-			+			CGAG + GCTC	360
40		L	N	G	K	E	Y	K	с к	: v	s	N	K	A	L	P	A	P	I	E	•
	361				-+-		 .	-+-			-+-			+ -			• • •			CCCA GGGT	420
45		ĸ	T	I	s	ĸ	A	ĸ	G () P	R	E	P	Q	v	Y	T	L	P	P	-
50	421				-+-			+-			-+-	• • • •	• • • •	+-			• • •			CTAT GATA	400
		s	R	D	E	L	T	K	N (2 V	S	L	T	С	L	V	K	G	F	Y	•
55	481				-+-			+-			-+-		• • • •	+	• • • •	• • • •	4			GACC CTGG	340
			_	_	_	_															

FIG.5B

_			GCC'	rcc	CGT	GCT	GGA	CTC	CGA	CGG	CTC	CTT	CTT	CCT	CTA	CAG	CAAC	CTC	ACC	GTO	GAC	600
5	541	TG	CGG	AGG	GCA	CGA	CCT	GAG	GCT	GCC	GAG	GAA(GAA	GGA	GAT	STC	STT	CGAC	TGG	CAC	CTG	
		T	P	P	v	L	D	S	D	G	s	F	F	L	Y	s	ĸ	L	T	v	D	-
10	601				-+-			+				+		• • •	-+-	•		+ .			CAC	660
	001	TT	CTC	GTC	CAC	CGT	CGT	ccc	CTT	GCA	GAA	GAG'	rac	GAG	GCA(CTA	CGT	ACTO	CGA	GAC	GTG	
4.5		ĸ	s	R	W	Q	Q	G	N	v	F	s	С	s	v	M	H	E	A	L	Н	•
15		AA	CAC	CTA	CAC	GCA	GAA	GAG	CCT	CTC	CCT	GTC:	rcc	GGG'	raa.	AGT	ACC	GATO	CAG	AA	AGTT	720
	661	TT	 GGT(GAT(·+· GTG	CGT	CTT	+ CTC	GGA	GAG	GGA	CAG	AGG	ccc	ATT'	TCA'	rgg	CTA	GTC	TT	CAA	120
20									L													•
		CA	GGA	CGA	CAC	CAA	AAC	CTT	AAT	TAA	AAC	GAT	CGT	TAC	GCG'	TAT	CAA	CGA	CATO	AG	CAC	700
	721	GT	CCT	 GCT	-+- GTG	GTT	TTG	GAA	TTA	ATT	TTG	+ CTA	GCA	ATG	- + - CGC.	ATA	GTT(GCT	GTAC	TC	AGTG	780
25									r													•
		AC	CCA	GTC	GGT	GAG	CTC	TAA	ACA	GAA.	AGT	TAC.	AGG	CCT	GGA	CTT	CAT	ccc	GGG:	CTC	GCAC	940
30	781	TG	 GGT	CAG	-+- CCA	CTC	GAG	TTA	TGT	CTT	TCA	+ ATG	TCC	GGA	CCT	GAA	GTA	GGG	CCC	AGA	CGTG	040
		T	Q						Q									P				•
				CCT	GAC	CTT	GTC	CAA	AAT	'GGA	CCA	GAC	CCT	GGC	TGT	ATA	CCA	GCA	GAT(CTT	AACC	900
35	841	GG	CTA	GGA	CTG	GAA	CAG	GTT	TTA	CCT	GGT	CTG	GGA	CCG	ACA	TAT	GGT	CGT	CTA	GAA'	TTGG	,,,,
		P	I	L	T	L	s	K	M	D	Q	T	L	A	v	Y	Q	Q	I	L	T	-
40			CAT	GCC	GTC	CCG	TAA	CGI	TAT	CCA	GAT	CTC	TAA	CGA	CCT	CGA	GAA	CCT	TCG	CGA	CCTG	960
	901	AG	GTA	CGG	CAG	GGC	ATI	GCA	ATA	GGT	CTA	GAG	ATT	GCT	GGA	GCT	CTT	GGA	AGC	GCT	GGAC	,,,,
4.5		s	M	P	s	R	N	V	I	Q	I	s	N	D	L	E	N	L	R	D	L	•
45		CT	GCA	CGT	GCI	GGC	:ATT	CTC	CAA	ATC	CTG	CCA	CCT	GCC	ATG	GGC	TTC	AGG	TCT'	TGA	GACT	1020
	961	GA	CGT	GCA	CGA	CCC	TAP	GAC	GTI	TAC	GAC	GGT	GGA	CGG	TAC	CCG	AAG	TCC	AGA	ACT	CTGA	1020
50		L	н	v	L	A	F	s	K	s	С	н	L	P	W	A	s	G	L	E	T	•
			GGA	CTC	TCI	'GGG	CGC	GGT	rcci	rgga	AGC	ATC	:CGG	TTA	CAG	CAC	CGA	AGT	TGT	TGC	TCTG	1080
	1021	GA	CCI	GAG	AG/	ACCO	GCC	CCZ	\GG/	ACCI	TCC	TAG	GCC	AAT	GTC	GTG	GCT	TCA	ACA	ACG	AGAC	1080
55		L	D	s	L	G	G	v	L	E	Α	s	G	Y	s	T	E	v	v	A	L	•

FIG.5C

TCCCGTCTGCAGGGTTCCCTTCAGGACATGCTTTGGCAGCTGGACCTGTCTCCGGGTTGT

AGGGCAGACGTCCCAAGGGAAGTCCTGTACGAAACCGTCGACCTGGACAGAGGCCCAACA

S R L Q G S L Q D M L W Q L D L S P G C

TAATGGATCC

1141 TAATGCTAGG

ATTACCTAGG

FIG.6A

Recombinant human metFc-OB (double stranded) DNA and amino acid sequence (SEO ID NOS: 3 and 4).

J				7 00			~~~															
	1				+		• • •	•	+		<i>.</i>	-+-			+						STCCI SAGGA	- 60
10			M	D	K	T	н	T	С	P	P	С	P	A	P	E	L	L	G	G	P	•
15	61			• • •	• • +			• • •	+		• • •	-+-			+	· • • ·		+			TGAG	120
13		s	v	F	L	F	P	P	K	P	K	D	T	L	М	I	s	R	T	P	E	•
20	121			• •	+				+		. .	+		· ·	-+-			+			GTAC + CATG	180
		V	T	С	V	V	V	D	V	S	Н	E	D	P	E	V	K	F	N	W	Y	•
25	181				+ -	• • • •	• • • •	4	+			+			-+-			+		• • •	CAGC + GTCG	240
		v	D	G	v	E	v	н	N	A	K	T	ĸ	P	R	E	E	Q	Y	N	s	-
30	241	• -			-+-		• • • •	• • • +				+			-+-		• • •	+			GGAG + CCTC	300
35		T	Y	R	v	v	s	v	L	T	v	L	н	Q	D	W	L	N	G	K	E	•
	301	• •			-+-			+			• • •	+			-+-			+			CAAA + GTTT	360
10		Y	ĸ	С	K	V	S	N	K	A	L	P	A	P	I	E	ĸ	T	I	s	K	-
15	361		• • •	• • •	-+-		• • •	+			• • •	+	• • •		-+-	•		+		. .	GCTG	420
		A	ĸ	G	Q	P	R	E	P	Q	v	Y	T	L	P	P	s	R	D	E	L	•
50	421	• • •			-+-			+	• • •		• • •	+	• • •		-+-			• • • •			GCC GCGG	480
		T	ĸ	N	Q	v	s	L	T	С	L	v	ĸ	G	F	Y	P	s	D	I	A	•
5	481		· ·		-+-			+	• • •			+			-+-			-+-	. .		CTG GAC	540
		v	E	W	E	s	N	G	0	P	E	N	N	Y	ĸ.	т	т	P	Þ	v	ī.	

FIG.6B

5	- 41					CTC	CTT	CTT	CCT	CTA	CAG	CAA	GCT(CAC	CGT	GGA	CAA	3AG(AG	i I G	CAG	600
5	, 241	CT	GAGGCTGCCGAGGAAGAAGGAGATGTCGTTCGAGTGGCACCTGTTCTCGTCCACCGTC																			
		D	s	D	G	s	F	F	L	Y	S	ĸ	L	T	v	D	K	S	R	W	Q	•
10			GGG	GAA	CGT	CTT	CTC.	ATG	CTC	CGT	GAT	GCA'	TGA	GGC	TCT	GCA	CAA	CAC	CTA(CAC	GCAG	660
	601	GT	ccc	CTT	GCA	GAA	GAG	TAC	GAG	GCA	CTA	CGT	ACT	CCG	AGA	CGT	GTT(GGT	GAT(GTG(CGTC	
		Q	G	N	v	F	s	С	s	V	M	H	E	A	L	Н	N	H	Y	T	Q	•
15																	TCA	GGAC	CGA	CAC	CAAA	720
	661	TT	CTC	GGA	GAG	GGA	CAG.	AGG	ccc.	ATT	TCA	TGG	CTA	GGT	CTT'	TCA	AGT				STTT	, 20
20		ĸ	s	L	s	L	s	P	G	K	v	P	I	Q	ĸ	v	Q	D	D	T	ĸ	•
	701		CTT.	AAT	TAA	AAC	GAT	CGT	TAC	GCG	TAT	CAA	CGA	CAT	CAG	TCA	CAC	CAC	STC	GT	GAGC	780
1 E	721	TG	GAA	TTA																	CTCG	
25		T	L	I	K	T	I	v	T	R	I	N	ם	I	s	H	T	Q	S	v	S	•
	781																		CT	GAC	CTTG	840
30	701	AG	TTA	TGT	CTT	TCA	ATG	TCC	GGA	CCT	GAA	GTA	GGG	CCC	AGA	CGT	GGG	CTAC	GGA(CTG	GAAC	
		s	K	Q	K	V	T	G	L	D	F	I	P	G	L	Н	P	I	L	T	L	•
35	841				-+-			+			• • •	+			-+-			+			CCGT	900
		AG	GTT	TTA	CCT	GGT	CTG	GGA	CCG	ACA	TAT	GGT	CGT	CTA	GAA	TTG	GAG	GTA(CGG	CAG	GGCA	
		S		М	_	Q	T	L		٧	_	-	Q	_	L		S	М	P	S	R	•
40	901				-+-			+		• • •		+••			-+-			• • +				960
		TT	GCA	ATA	.GGT	CTA	GAG	ATT	GCT	GGA	GCT										CCGT	
45		•	V	_	-		_	N	_								L				Α	•
	961				-+-			+				+	• • •		-+-	• • •	• • •	+	• • •		GGGC	1020
		AA	GAG	GTT	TAG	GAC	GGT	GGA	CGG	TAC	CCG	AAG	TCC	AGA	ACT						cccg	
50		F	-	K	S	С		L	P	•	Α	_	G	L	E	T.	L	D 	s 	L	G 	•
	1021				-+-			+				+	· · ·		-+-	•	• • •	+	• • •	• • •		1080
55		CC																			CCCA	
		G	V	L	E	Α	S	G	Y	S	T	E	V	V	Α	L	S	R	L	Q	G	•

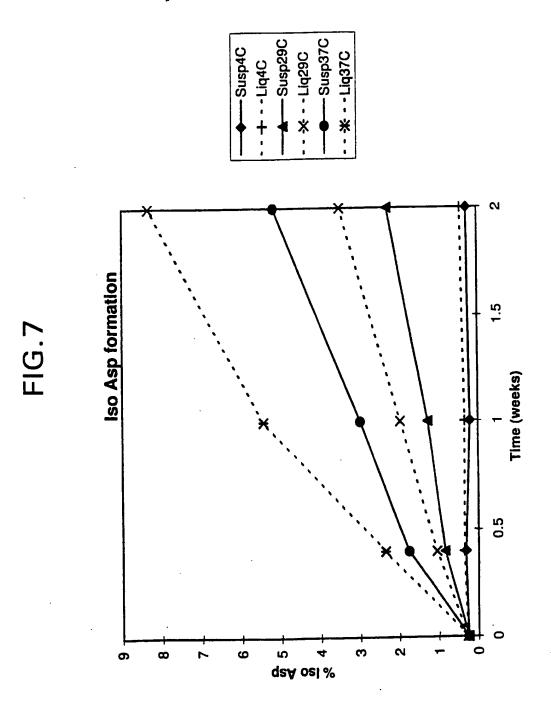
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11/12

FIG.6C

5	1081		•••		-+-		•••	+	•••			+			-+-		TTAATGGATCC AATTACCTAGG	1135
		s	L	Q	D	M	L	W	Q	L	D	L	s	P	G	С	•	-

12/12



ti ational Application No PCT/US 98/07828

A. CLASSI IPC 6	FICATION OF SUBJECT MATTER A61K38/22		
According to	nternational Patent Classification (IPC) or to both national classification	tion and IPC	
	SEARCHED		
	cumentation searched (classification system followed by classification A61K C07K	n symbols)	
	ion searched other than minimum documentation to the extent that su	·	rched
Electronic da	ata base consulted during the international search (name of data bas	e and, where practical, search terms used)	
C. DOCUME	ENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the rele	vant passages	Relevant to claim No.
X	WO 97 00319 A (SMITHKLINE BEECHAM; BROWNE MICHAEL JOSEPH (GB); CHAP CONRAD) 3 January 1997 see page 7, line 33 - line 35 see page 8, line 16 - line 22; clexamples	MAN	1-13
Ρ,Χ	WO 97 24440 A (GENENTECH INC ;SAU FREDERIC J DE (US); LEVIN NANCY (VANDL) 10 July 1997 see claims; examples		10-13
A	WO 96 23518 A (LILLY CO ELI ;BASI MARGARET B (US); DIMARCHI RICHARD H) 8 August 1996 see page 25, line 10 - line 19	NSKI D (US);	1-9
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X Furth	ner documents are listed in the continuation of box C.	X Patent family members are listed in	n annex.
* Special ca	tegories of cited documents :	"T" later document published after the inten	national filing date
consid	ant defining the general state of the art which is not lered to be of particular relevance	or priority date and not in conflict with a cited to understand the principle or the invention "X" document of particular relevance; the cl	the application but ory underlying the
filing d	ate ont which may throw doubts on priority claim(s) or	cannot be considered novel or cannot involve an inventive step when the doc	be considered to
which	to site of the entertaint the marking disease of amothers	"Y" document of particular relevance; the cl cannot be considered to involve an inv	
"O" docume other r	ent referring to an oral disclosure, use, exhibition or means	document is combined with one or more ments, such combination being obvious	re other such docu-
	ent published prior to the international filing date but nan the priority date claimed	in the art. "&" document member of the same patent f	amily
Date of the	actual completion of theirternational search	Date of mailing of the international sear	ch report
2	7 August 1998	03/09/1998	
Name and n	nailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2	Authorized officer	
	NL - 2280 HV Rijswljk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Fuhr, C	

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Ir ational Application No PCT/US 98/07828

C (Continu	INTO DOCUMENTS CONSIDERED TO BE RELEVANT	101/03 90/0/020
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
	WO 98 28427 A (AMGEN INC) 2 July 1998 see claims; examples	10-13
		
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international application No.

PCT/US 98/07828

Box i Observations where certain claims were found unsearchable (C ntinuati n of item 1 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Remark: Although claims 10-13 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box il Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invitepayment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

Information on patent family members

Patent document cited in search repor	t	Publication date		atent : ms:
WO 9700319	Α	03-01-1997	AU EP	60 0
WO 9724440	Α	10-07-1997	AU	- · ·
WO 9623518	Α ·	08-08-1996	US AU CA EP	55 4 24 03
W0 9828427	A	02-07-1998	NONE	